

Göttingen Minipigs Genetics: Breed history, breeding strategies and genome analyses

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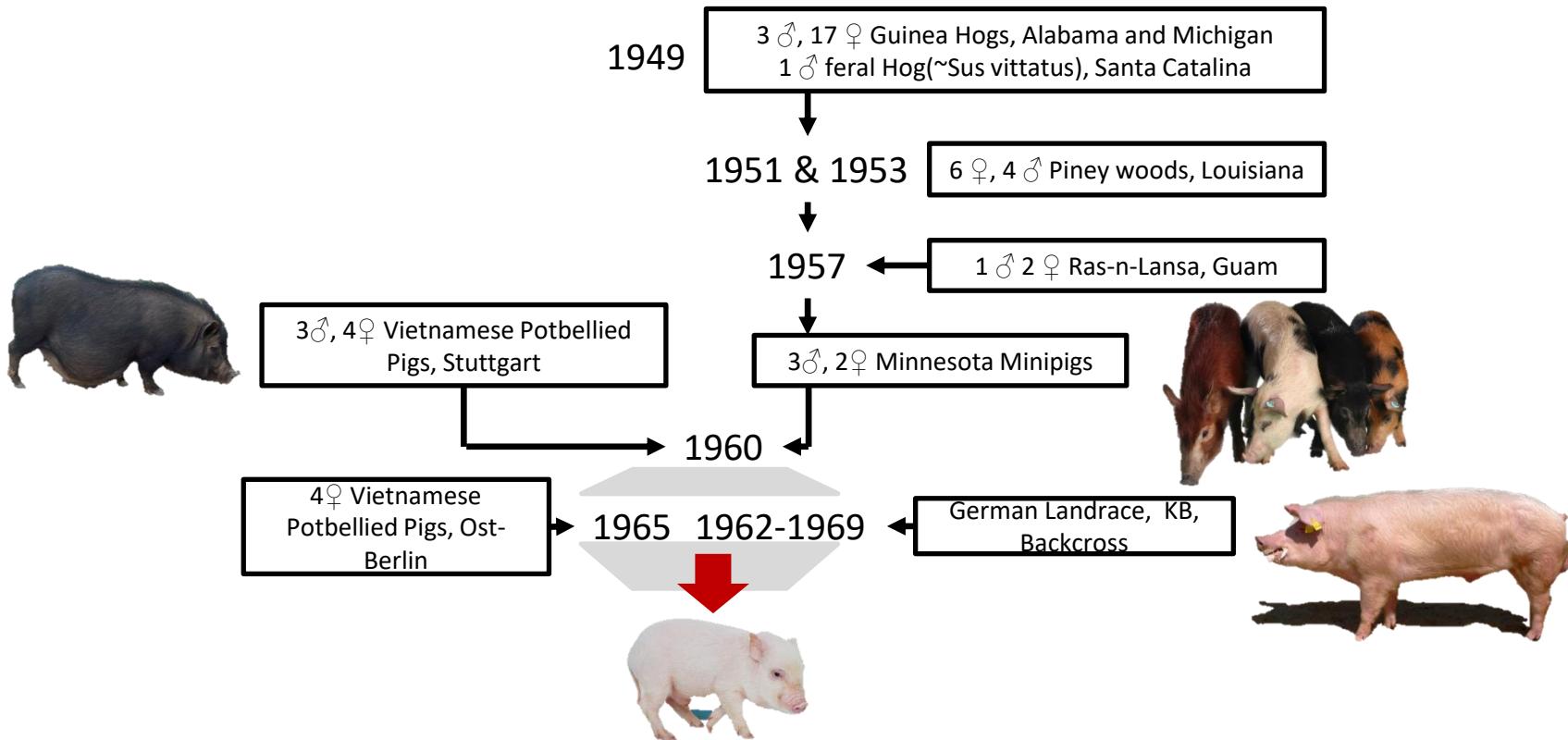


- History
- Breeding Program of the GMP
- World wide breeding structures
- Population Genetics
- Next Generation Sequencing (NGS)
- Assessing breed integrity of Göttingen Minipigs



History

Brief overview



The GMP today





First years

1960 bis 1969

Growing demand for animal model species

- Physiologically close to humans
- small
- But sufficiently large for surgeries
- uniform
- Moderate temperament
- Easy to house and handle



Prof. Fritz Haring
(1907 – 1990)

First years



1960: Import of first animals

- 3♂ und 2♀ Minnesota Minipigs (Hormel Institute, Austin, USA)
- 3♂ und 4♀ Vietnamese potbellied pigs (Wilhelma Zoo, Stuttgart)
→ heterogeneous coat colours

1965: Import of additional animals

- 4♀ Vietnamese potbellied pigs (Zoo Friedrichsfelde, East Berlin)
→ reduced size and white coat colour

1965 – 1969: Development of pure white-coated lines

- Artificial insemination with landrace
 - dominant white
 - lean
 - BUT larger

At Friedland



Minnesota Minipig (33%)



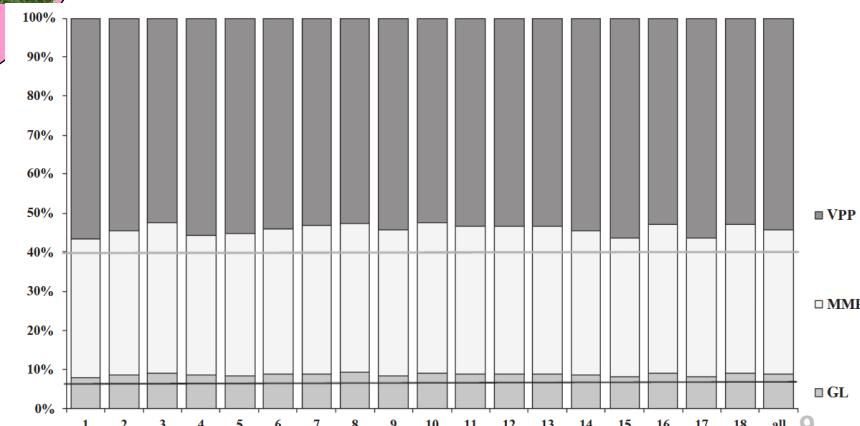
Vietnamese Potbellied pig (60%)



German Landrace (7%)



Gaerke et al. 2014



At Friedland



F₁-sow (Landrasse x Vietnamese)
x Minnesota boar

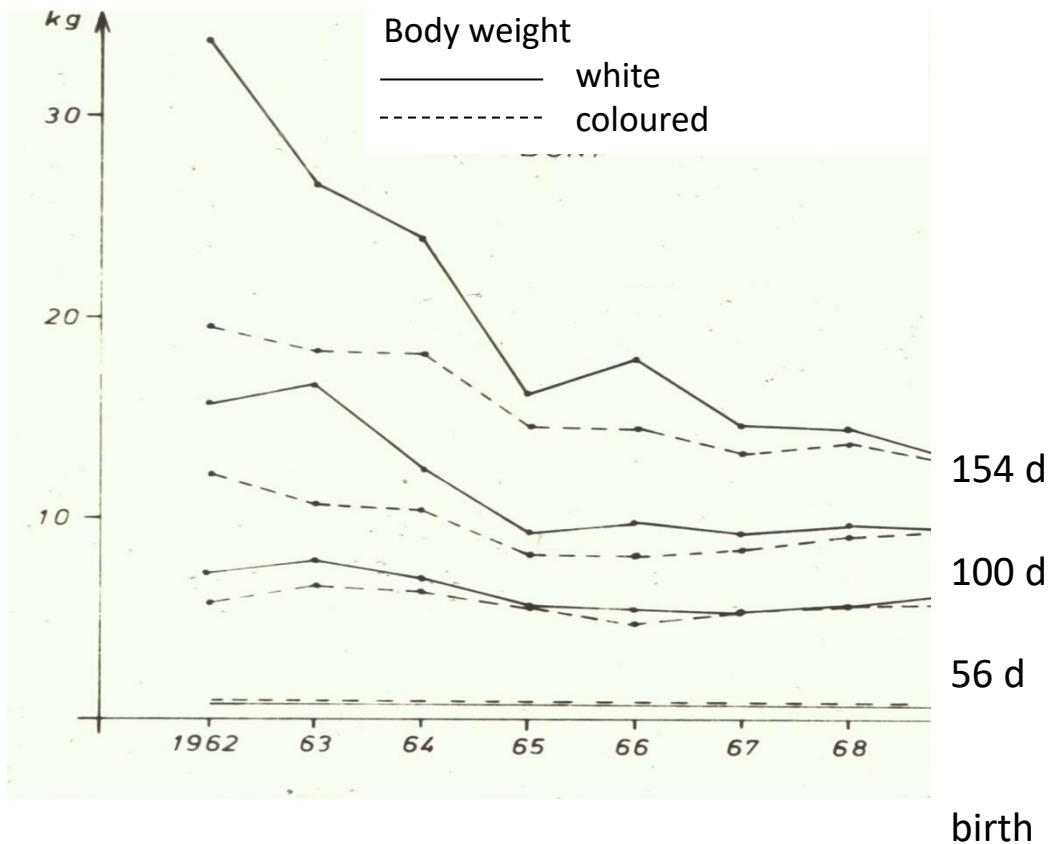
At Friedland



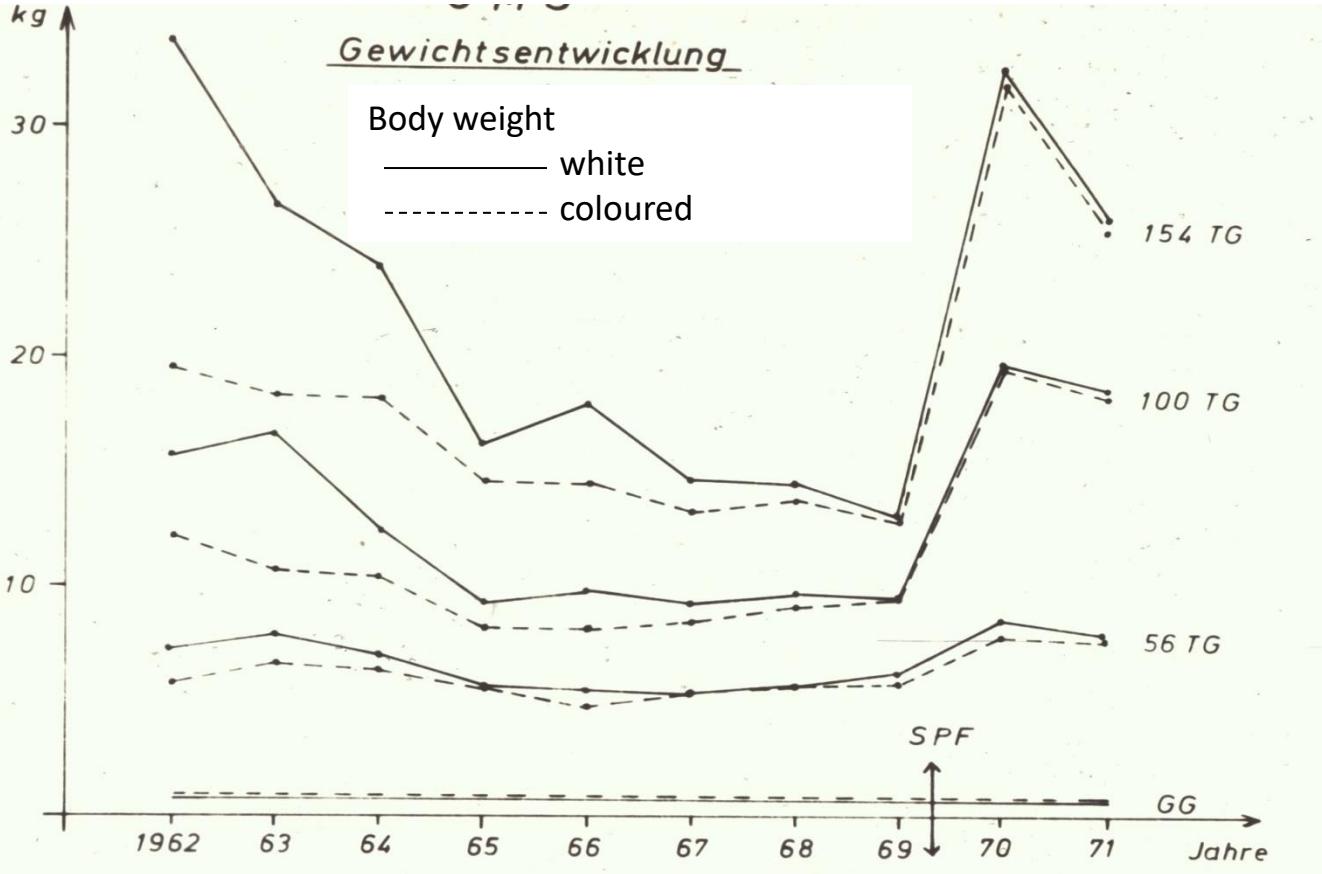
Low-input farming at Friedland led to

- high losses
- growth reduction due to hygienic problems
- lack of recording of phenotypes (5-month weight)

From Friedland to Relliehausen



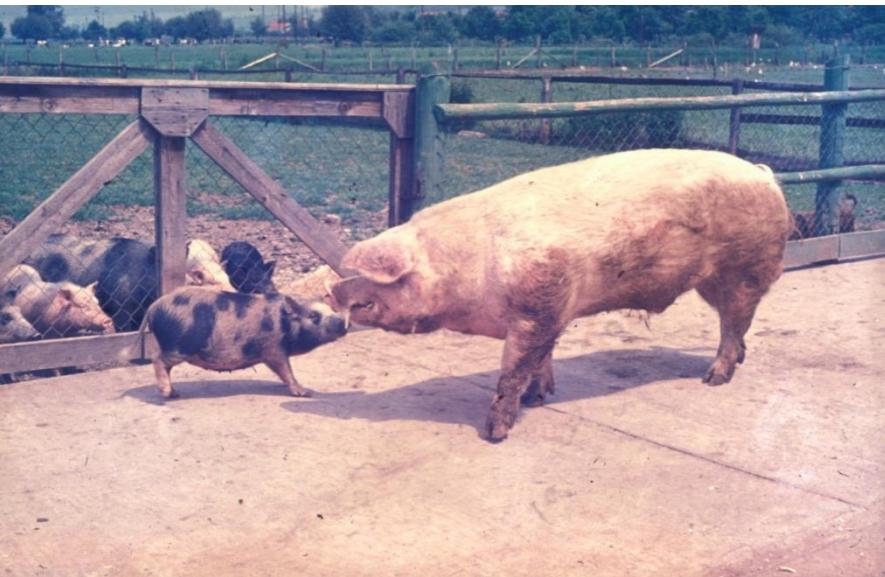
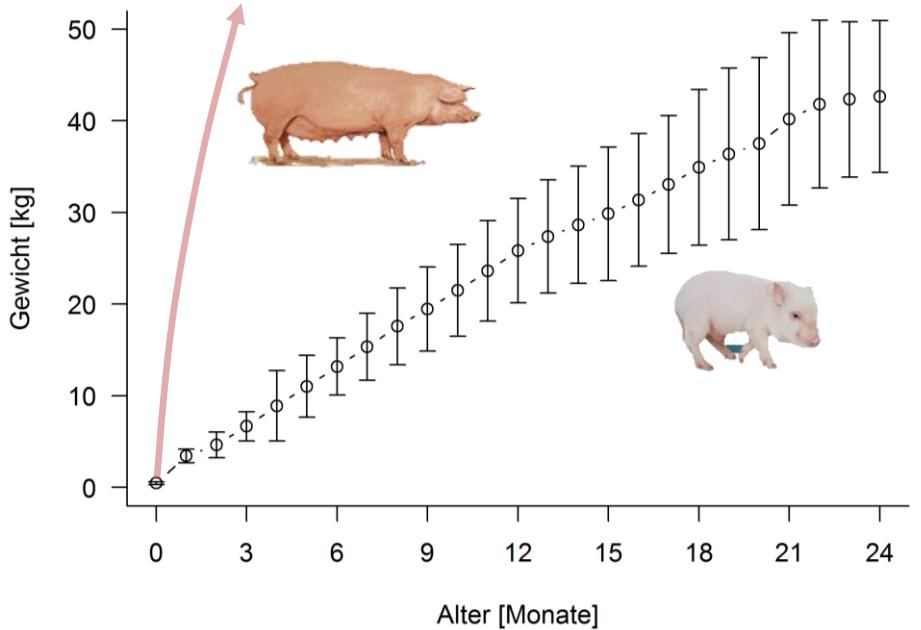
From Friedland to Relliehausen





Breeding Program of the GMP

Growth of the GMP



Breeding scheme



- Breeding goals
 - Weight
 - Relative Weight Reduction
 - Reproduction
 - Piglets born alive

Breeding scheme

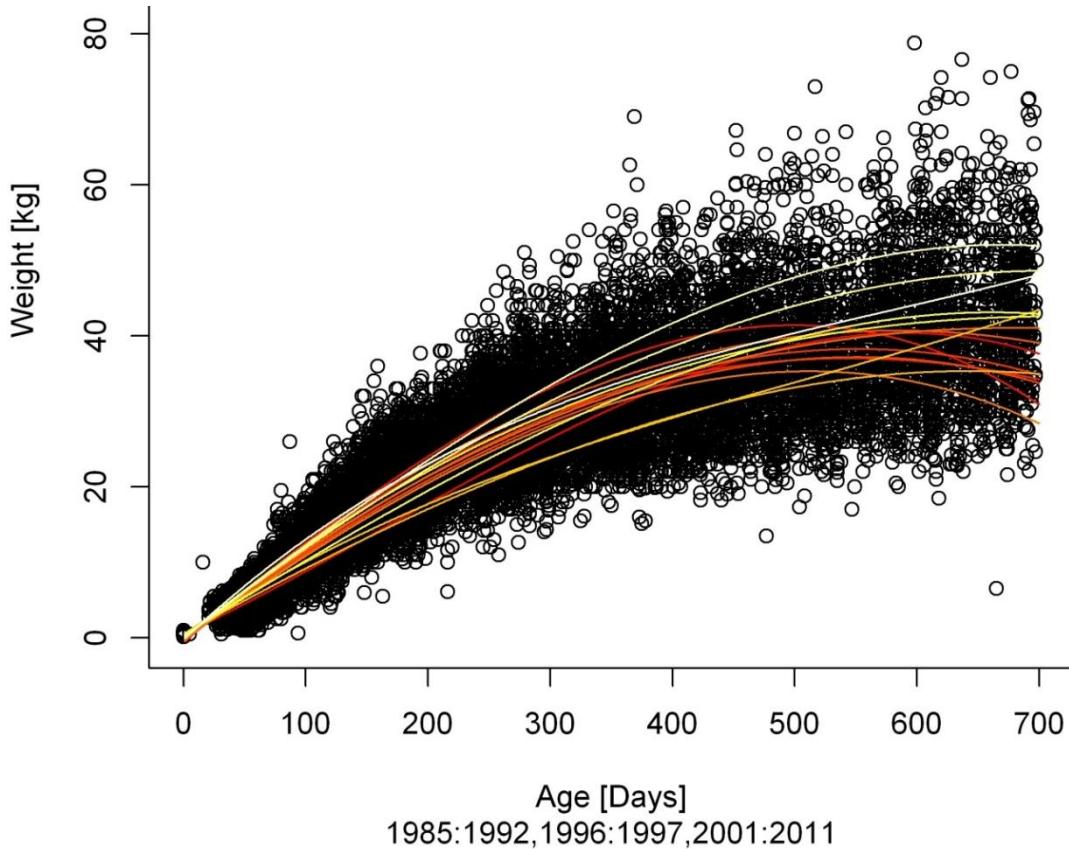


- Pedigree based breeding value estimation
- PostgreSQL database
- ongoing phenotyping
 - Weight
 - Piglets born alive and weaned
 - No. Of teats
 - Anomalies
- Optimum Genetic Contribution

Relative weight reduction



1. step: estimation of a growth curve



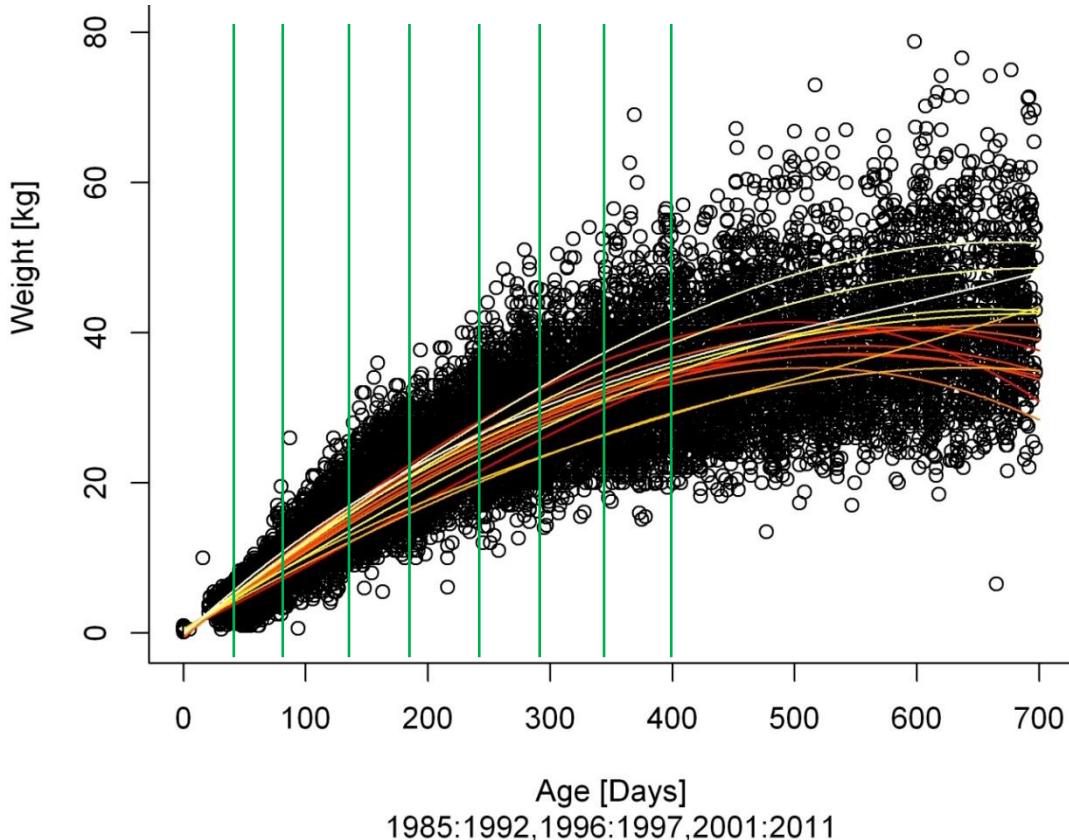
$$\text{weight} = \text{age} + \text{age}^2 + \text{age}^3 + \text{age}^4$$

1985:1992,1996:1997,2001:2011

Relative weight reduction



1. Comparison of 8 timepoints



$$\Delta = E(\text{gew}) - \text{gew}$$

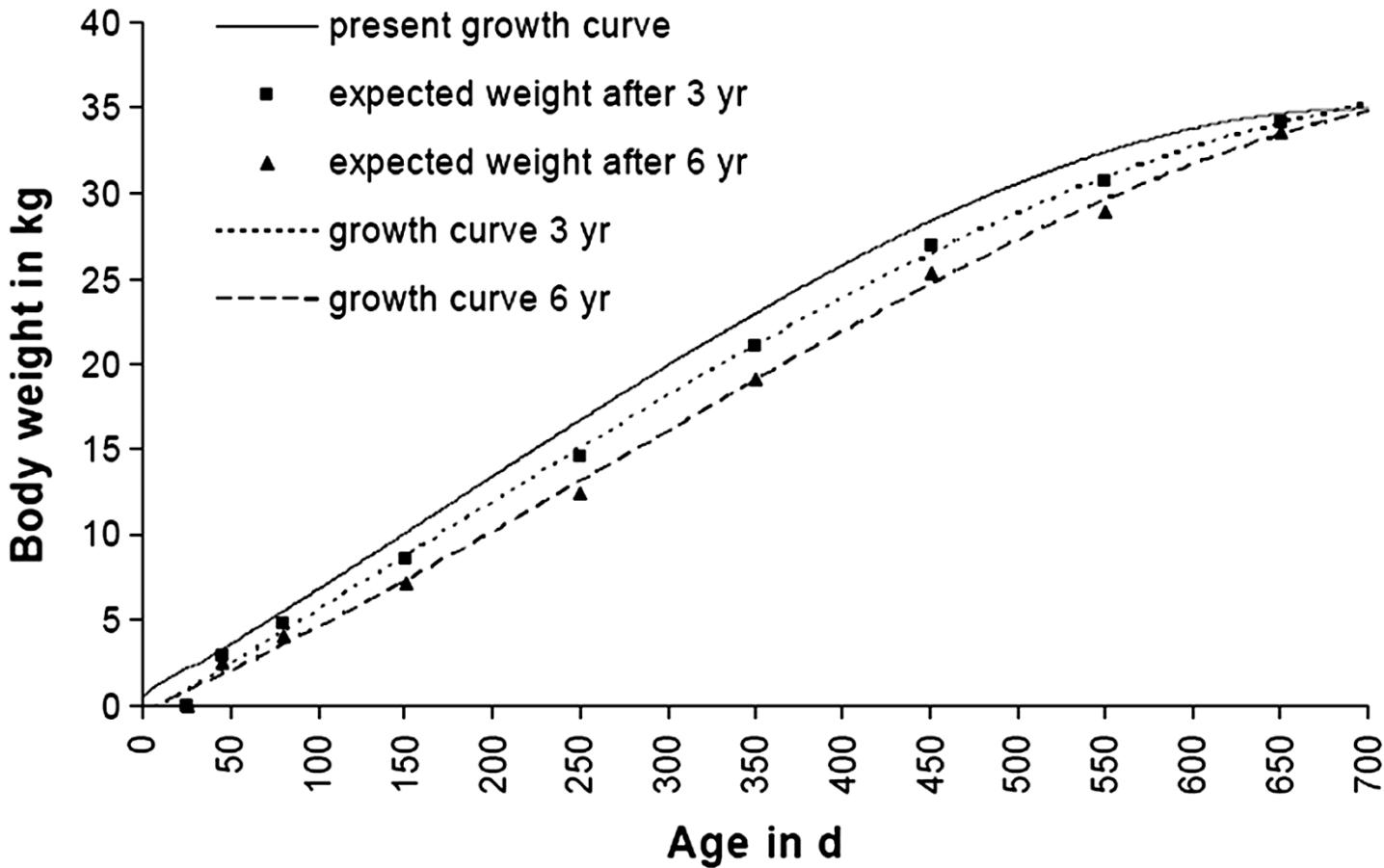


Relative weight reduction

- weighting of the differences to an index breeding value

	weighting factor	age class [days]
rwr1	0.00	less than 61
rwr2	0.31	61 – 100
rwr3	3.09	101 – 150
rwr4	2.62	151 – 201
rwr5	1.12	201 – 250
rwr6	0.73	251 – 300
rwr7	0.31	301 – 350
rwr8	0.20	351 – 400

Relative weight reduction



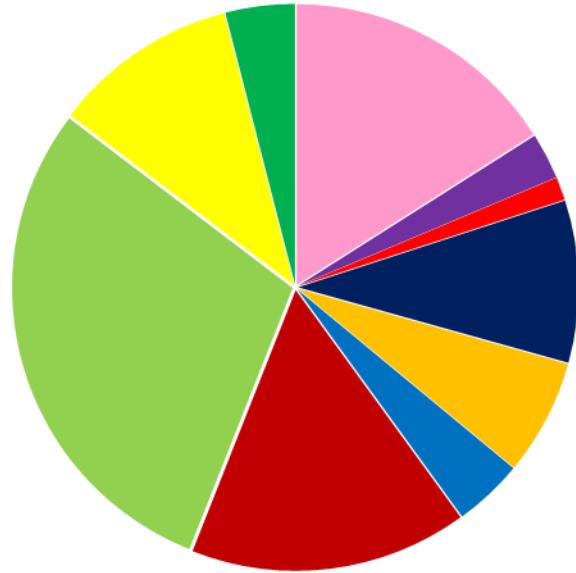
Optimum genetic contribution



10 Founder

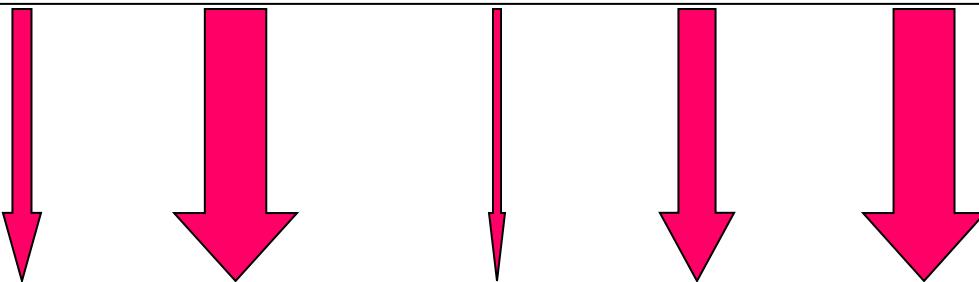


Optimal contribution
(minimum inbreeding)



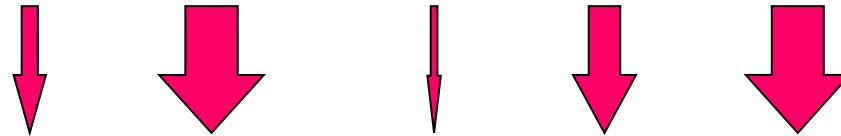
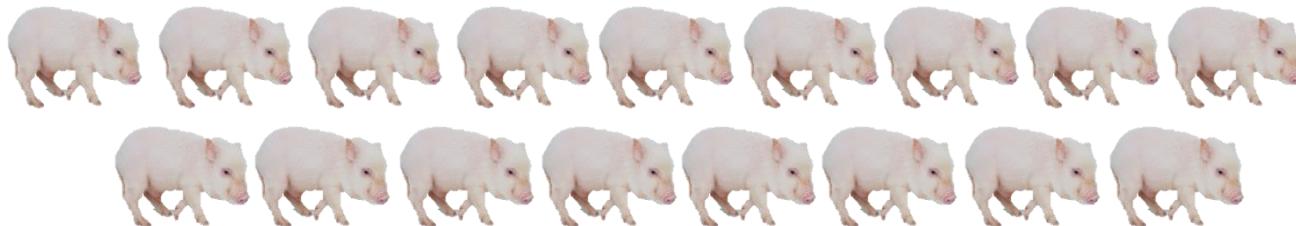
real contributions
(more than unavoidable inbreeding)²²

Optimum genetic contribution

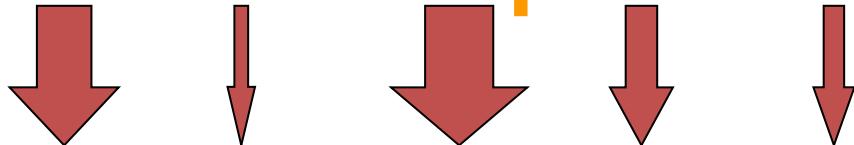


Current Population

Optimum genetic contribution



Current Population



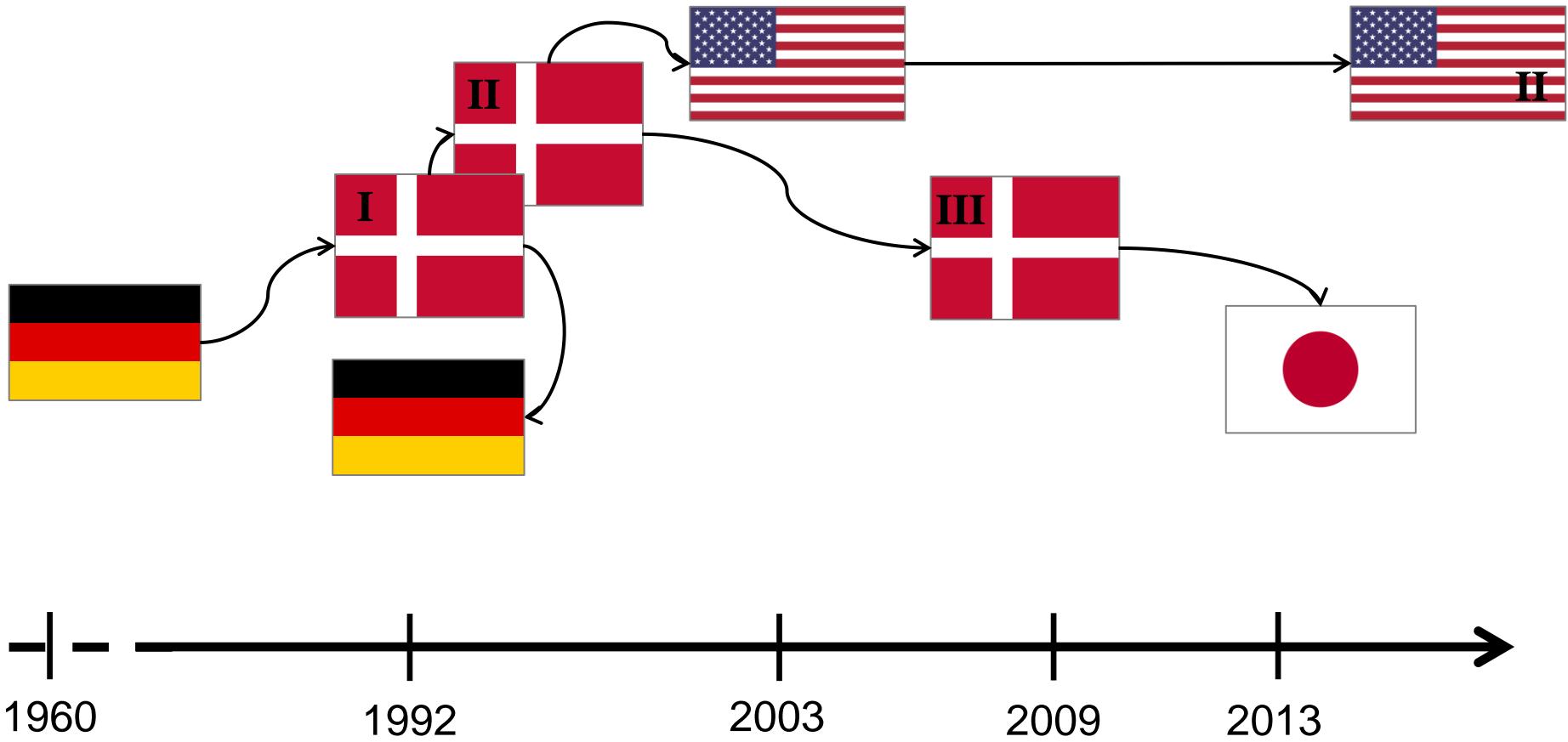
Optimum Contributions



World wide breeding structures



History of colonies

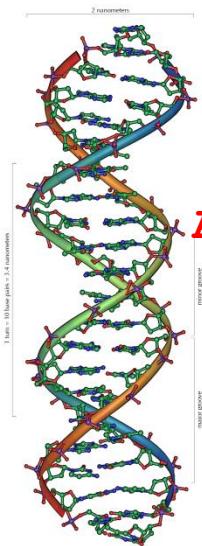


1960 1992 2003 2009 2013

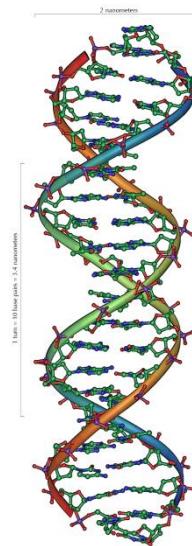


Population Genetics

Allele (Gene) frequency



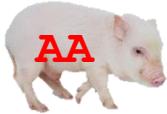
A Major allele in GMP



G Major allele in VPP

SNP [`Snipp`] = Single nucleotide polymorphism

Allele (Gene) frequency



$$p(A) = \frac{7}{10} = 0.7$$

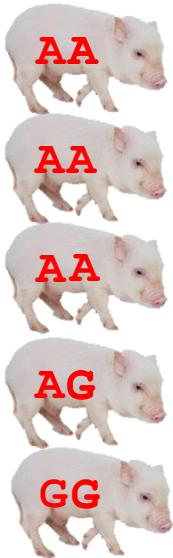
$$p(A) = \frac{3}{10} = 0.3$$

Allele frequency changes



- Genetic Drift Random transition of alleles to the next generation
- Selection „Fitness“ of an allele determines probability of transition
- Mutation Ancestral alleles mutate into alternative state with low probability
- Migration Allele exchange by introduction of new animals into population

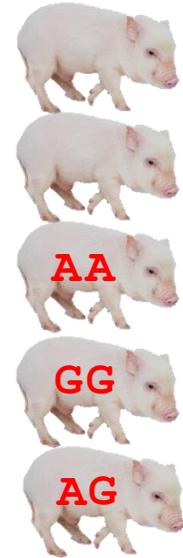
Drift



Generation t

$$p(A) = \frac{7}{10} = 0.7$$

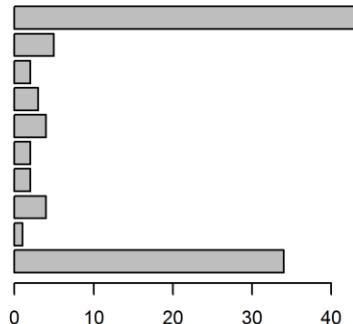
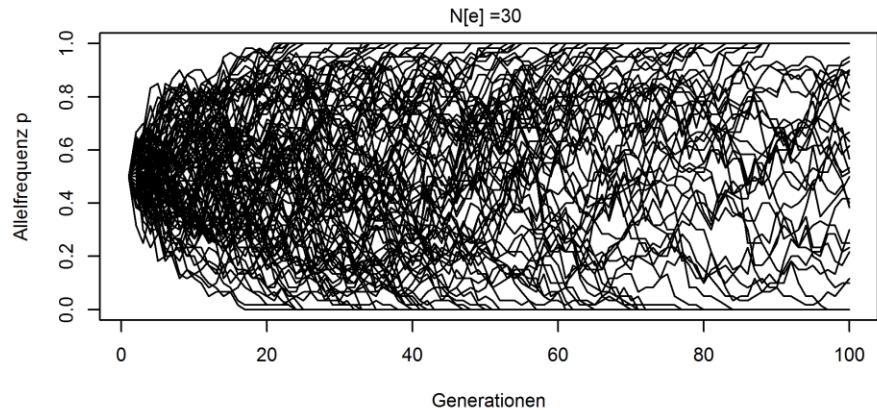
Drift is a process of random sampling with a binomial probability (in biallelic loci)



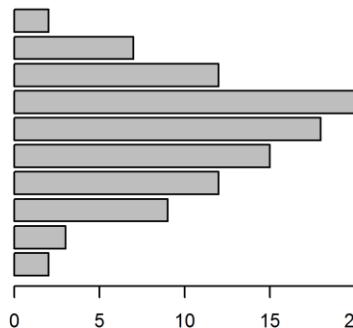
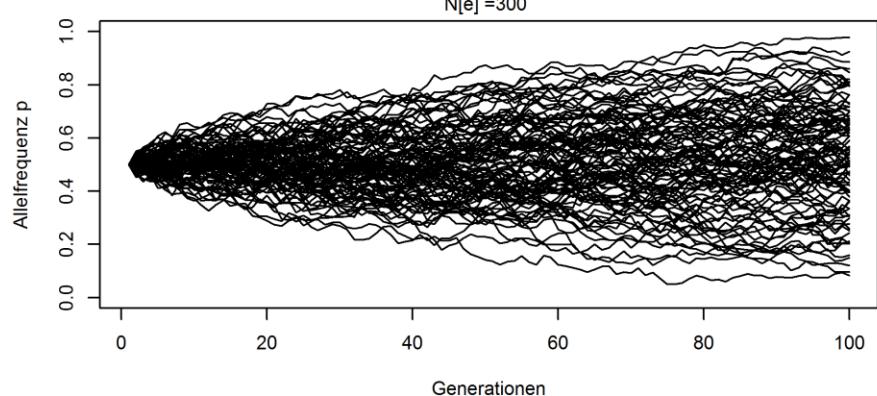
Generation $t + 1$

$$p(A) = \frac{4}{10} = 0.4$$

Drift



$$p(A) = 0.5$$
$$Ne = 30$$

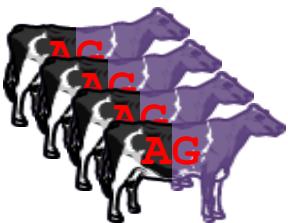


$$p(A) = 0.5$$
$$Ne = 300$$

Selection



1



$1 - \frac{1}{2}S$



$1 - S$

*Selection coefficient
 $S = 0.5$*

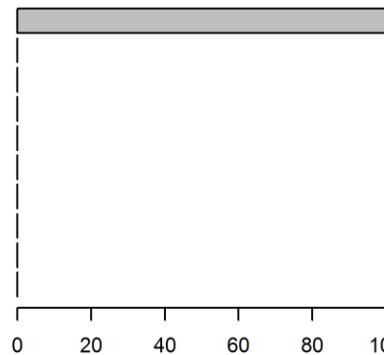
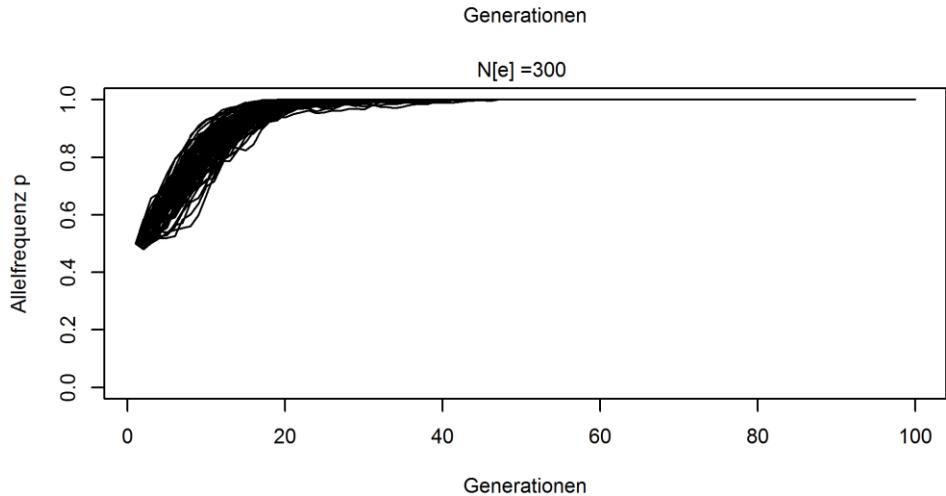
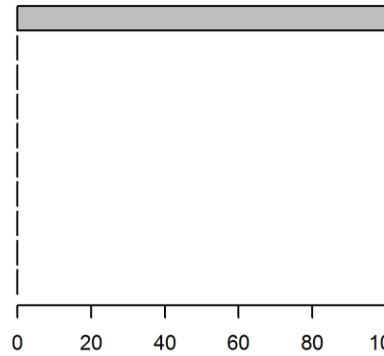
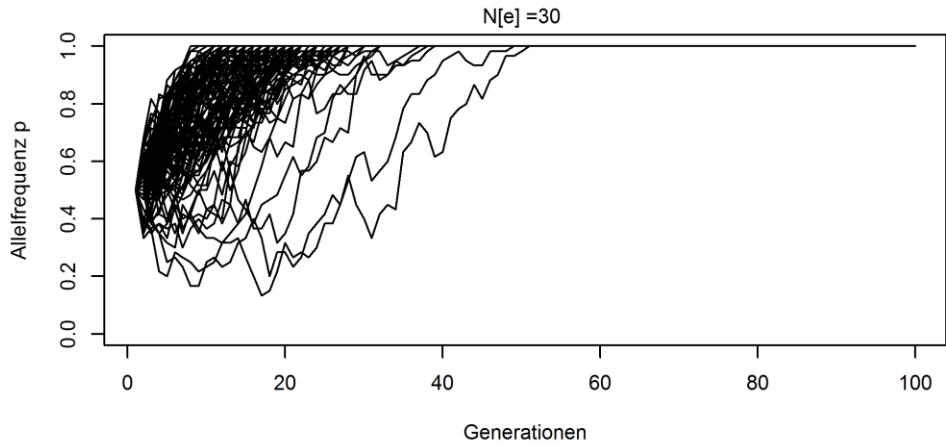
$$p_{A0} = 0.5$$

Gametic contribution
(without dominance)



$$p_{A1} = \frac{11}{18} = 0.611$$

Selection





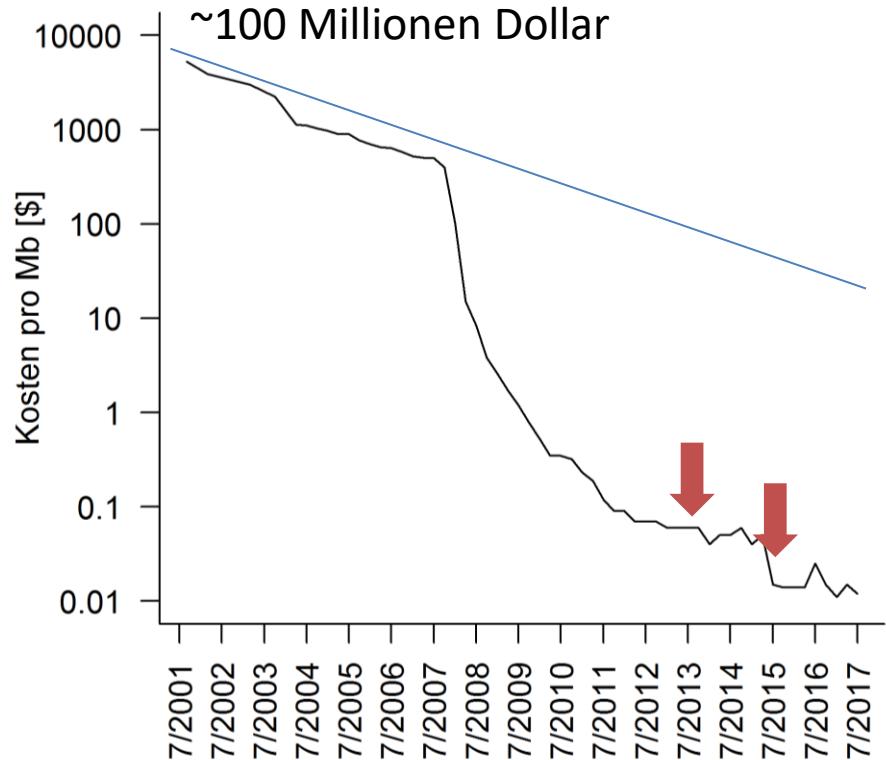
Next Generation Sequencing (NGS)

Whole-genome Shot-Gun Sequencing (WGS)

Next-Generation-Sequencing



- Whole genome represented
- Marker density to a multiple higher than for arrays
- Causal variation (theoretically) contained
- No array ascertainment bias
- Discovery of unknown variation





Reference genome



<http://www.nature.com>



1. Burrows-Wheeler-Alignment (BWA, Li und Durbin, 2009)



Reference genome



1. Sorting by chromosomal position with Samtools (Li et al., 2009)
2. Marking duplicates with Picard (Picard, 2009)
3. Indexing

→ **SAM/ BAM (Sequence Alignment/ Map - Format)**

→ **BAI Bam-Index**



Reference Genome

ACCCGTAGGGTTCATGGCATTGGTAA

ACCCCTA

CCCCTAG

CCGTAGG

CGTAGGG

CG

heterozygous

SNP

GCATTGA
CATTGAT
ATTGATA
TTGATAA

AA

homozygous

SNP



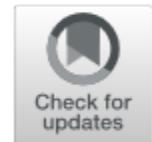
Reimer et al. *BMC Genomics* (2020) 21:308
<https://doi.org/10.1186/s12864-020-6590-4>

BMC Genomics

RESEARCH ARTICLE

Open Access

Assessing breed integrity of Göttingen Minipigs



Christian Reimer^{1,2*} , Ngoc-Thuy Ha^{1,2}, Ahmad Reza Sharifi^{1,2}, Johannes Geibel^{1,2}, Lars Friis Mikkelsen³, Martin Schlather^{2,4}, Steffen Weigend^{2,5} and Henner Simianer^{1,2}

Aim



- GMP colonies are genetically isolated, up to ~30 years
- Are they drifting apart from each other?
- Does breeding ensure selection for the same traits in all colonies?
- How much unique diversity is contained in each colony?
 - Risk of loosing a colony due to pests/ inbreeding/ fire?

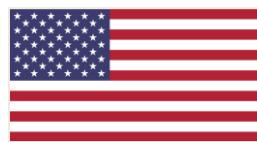
Materials & Methods - Sampling



DA2 1



DA2 2



NR 1



NR 2



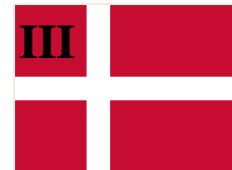
NI 1 NI 2



RE 1



RE 2



DA3 1

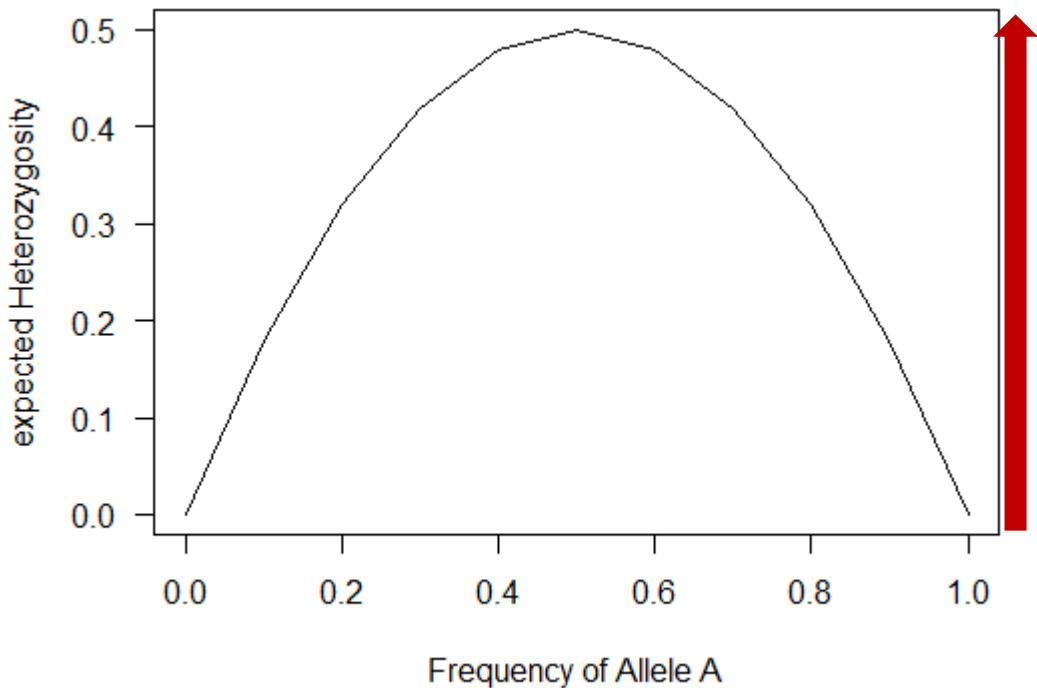


DA3 2



Expected Heterozygosity

- A measure of genetic diversity

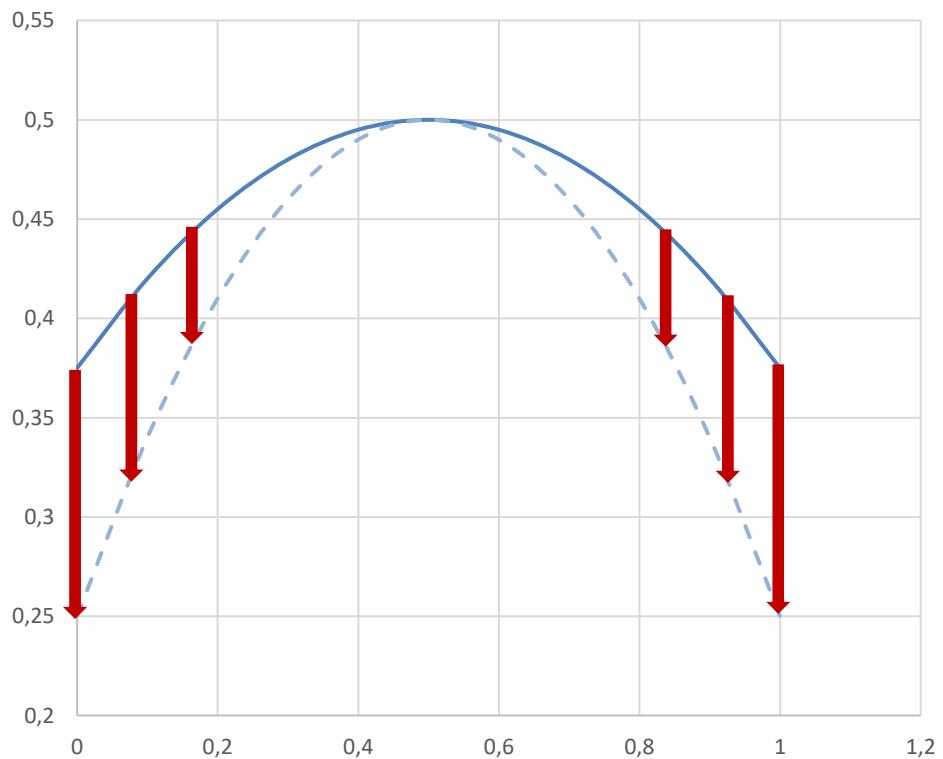


	$p(A)$	$p(a)$
$p(A)$	$p(A)^2$	$p(A)*p(a)$
$p(a)$	$p(A)*p(a)$	$p(a)^2$

Diversity

$$expHet = 2 * p(A) * (1 - p(A))$$

Wahlund Effect



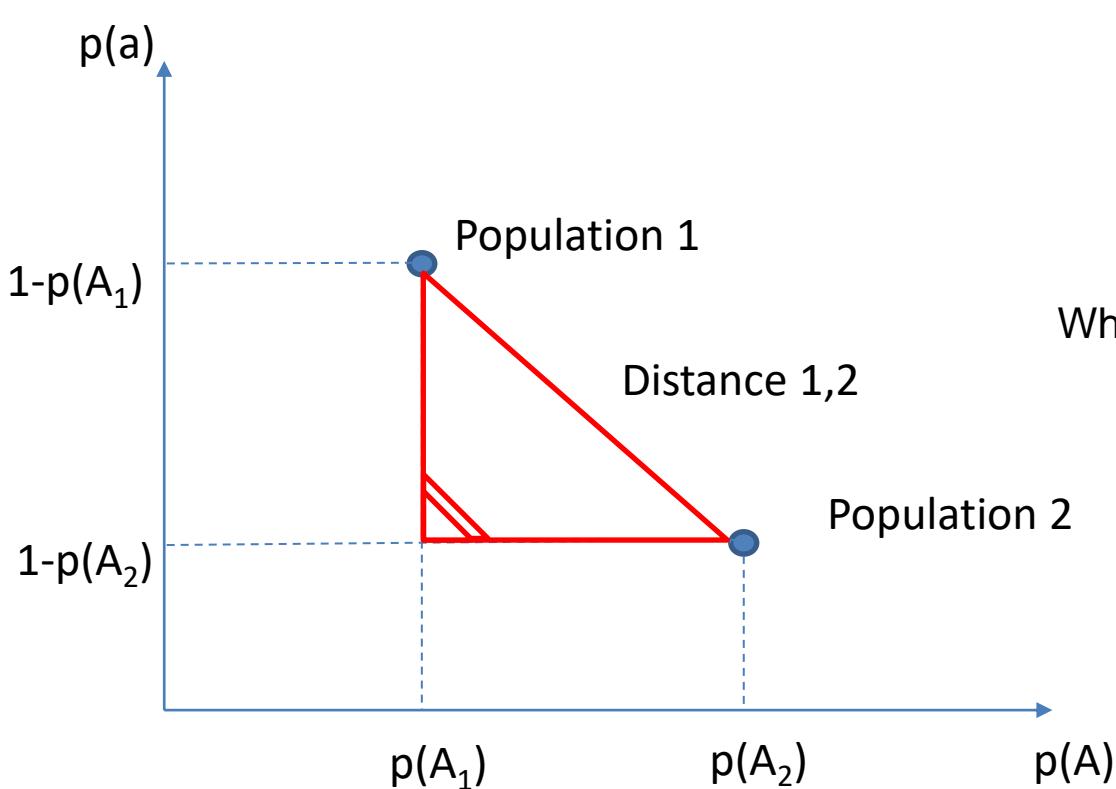
$$p(A)_{pop1} = 0.5$$
$$p(A)_{pop2} = x$$

— $expHet(\overline{p(A)})$

- - - $\frac{expHet(p(A)_{pop1}) + expHet(p(A)_{pop2})}{2}$

Decrease in
Heterozygosity due
to stratification
→ F_{ST}

Genetic distance



$$D = \sqrt{\sum_{i=1}^k (x_i - y_i)^2}$$

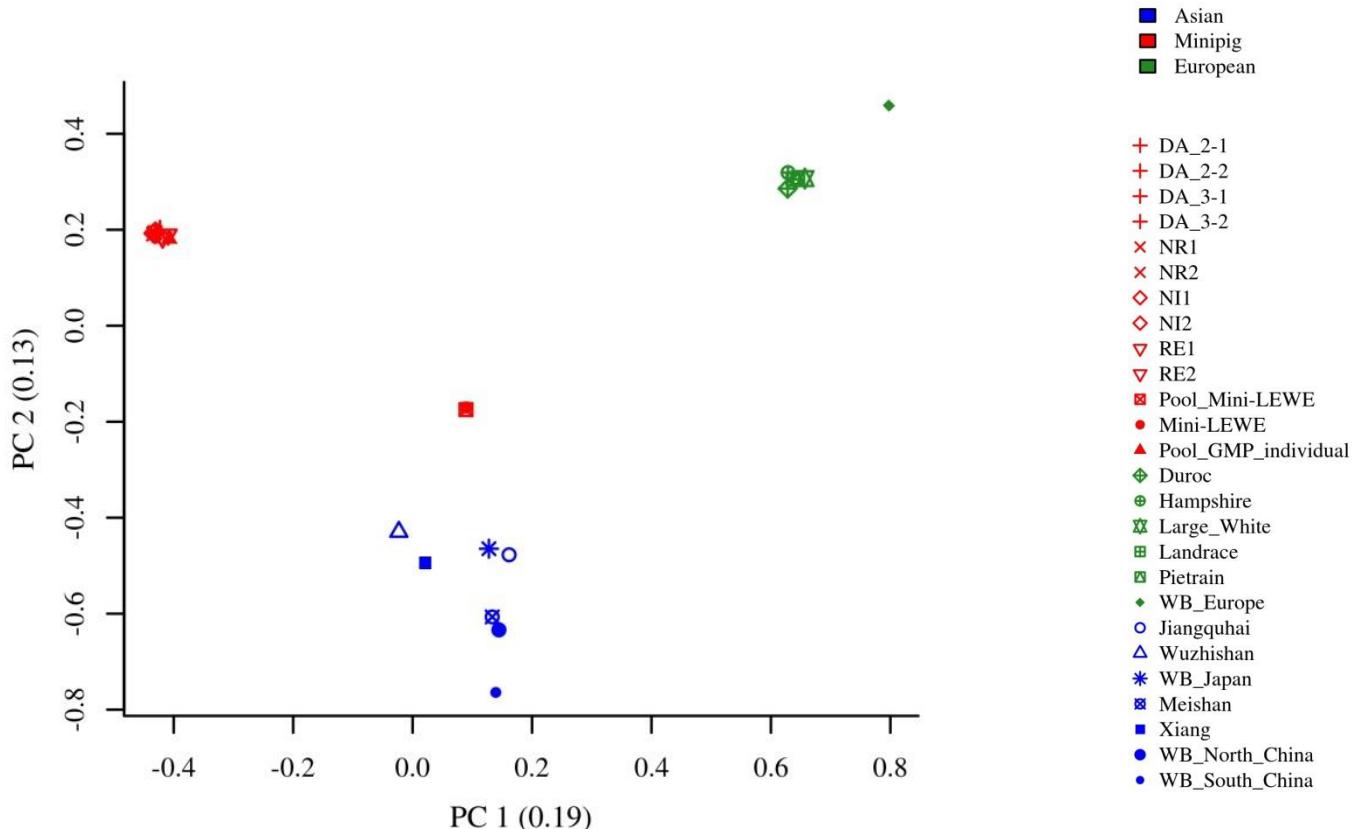
Where k – number of loci;
 x_i and y_i allele frequencies
at locus k of pop. i and j

Average Fst and Distances

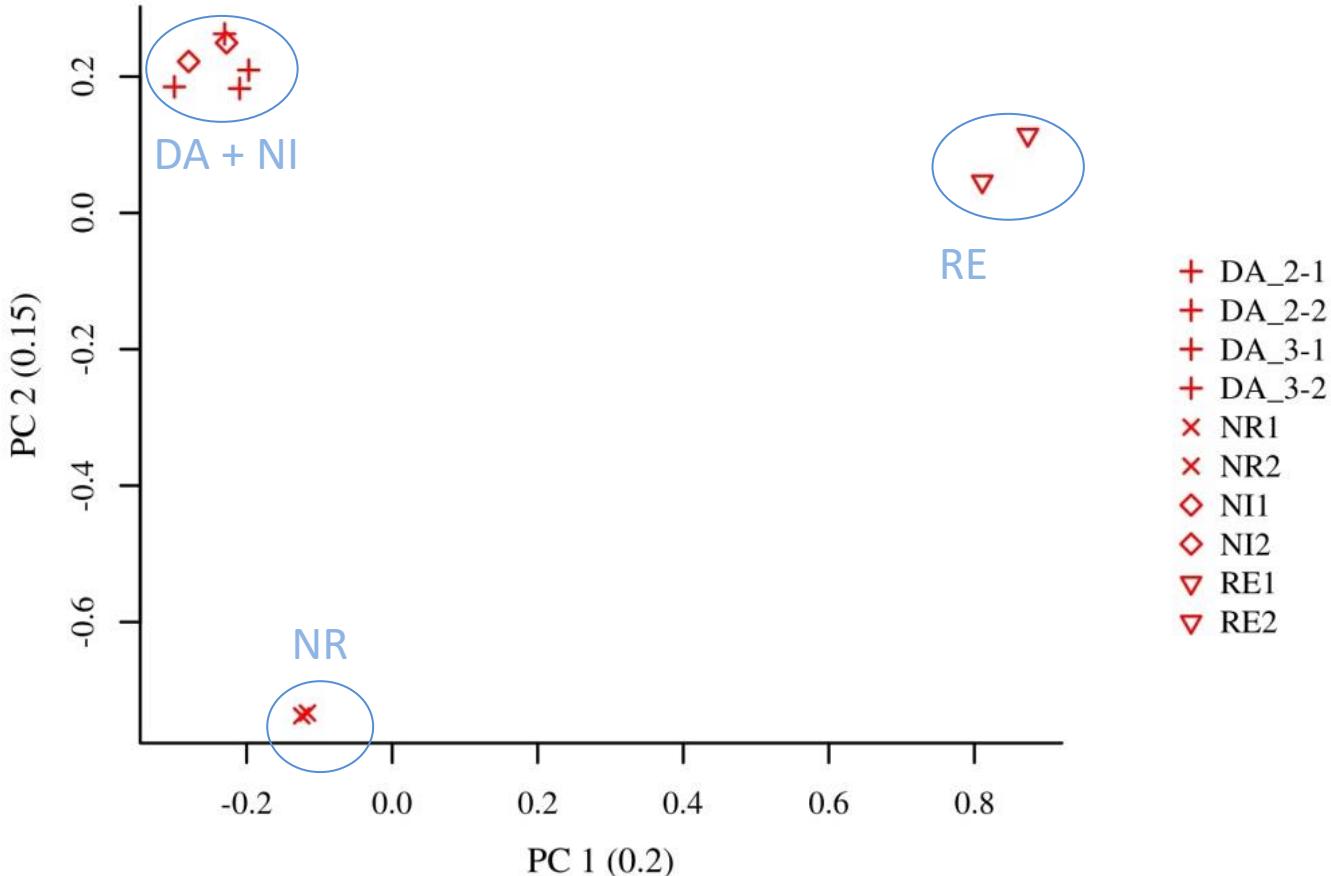


		GMP	Asian	European
FST	GMP	0.066	0.262	0.307
D _R	Asian		0.271	0.327
	European			0.163
D _R	GMP	0.113	0.364	0.408
D _R	Asian		0.372	0.428
	European			0.246

Principal component analysis



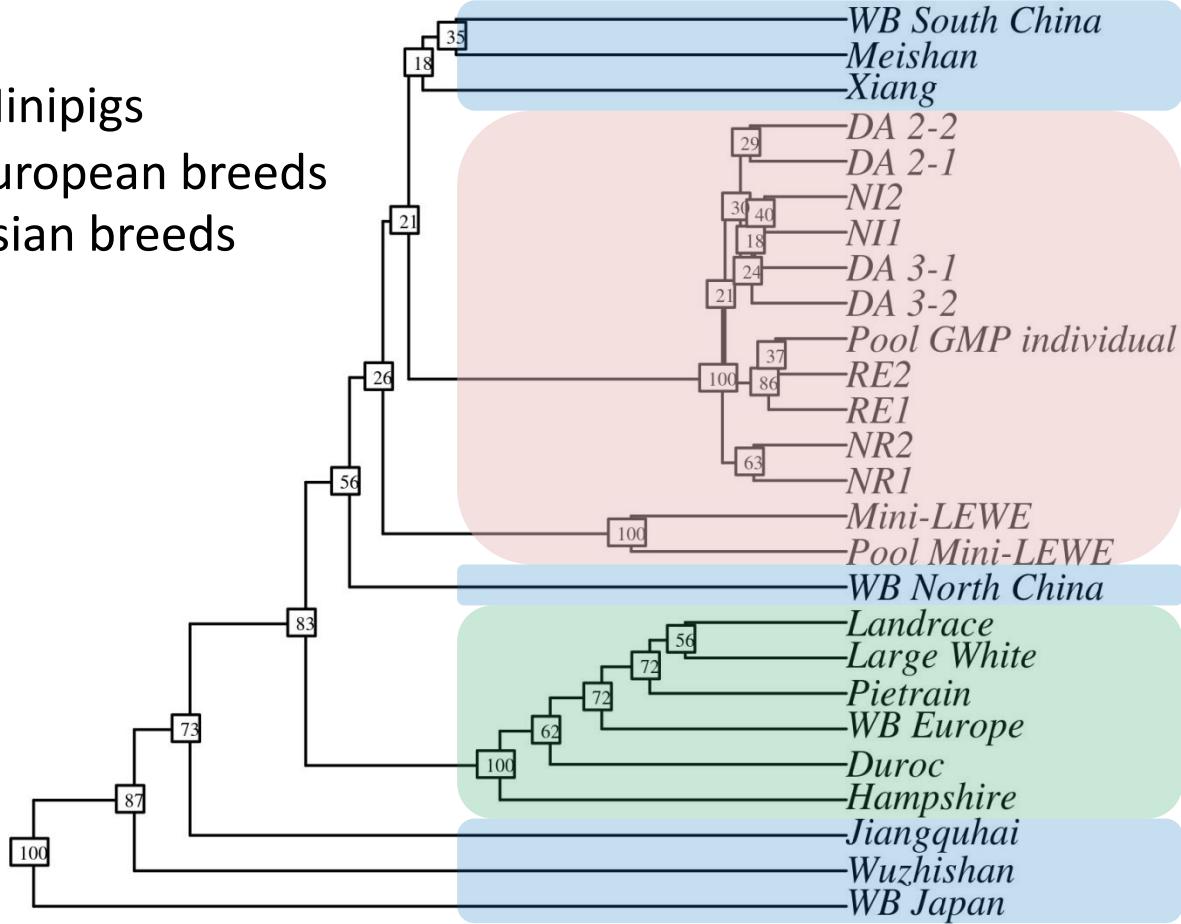
Principal component analysis



Phylogenie



Minipigs
European breeds
Asian breeds



Diversity within colonies



Expected Heterozygosity estimated from the virtual union of both unit pools

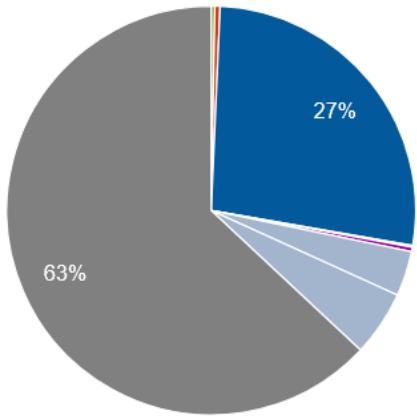
	RE	DA2	DA3	NR	NI
H_{exp}	0.298	0.292	0.294	0.285	0.295
SD	0.175	0.175	0.178	0.181	0.176
Nloci [M]	16.498	16.498	16.498	16.499	16.499
NNA	260	452	346	441	231

Private alleles

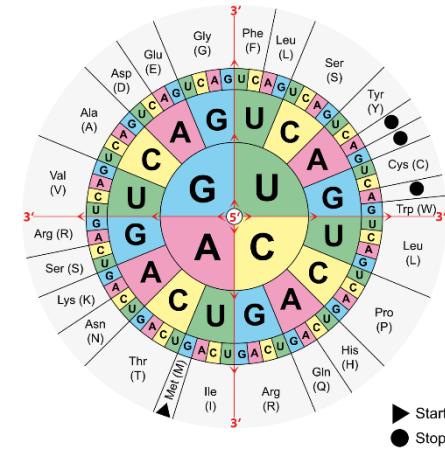


	No of private loci
RE	555'591
DA2	163'853
DA3	134'158
NR	192'896
NI	156'502

Ensembl variant effect predictor



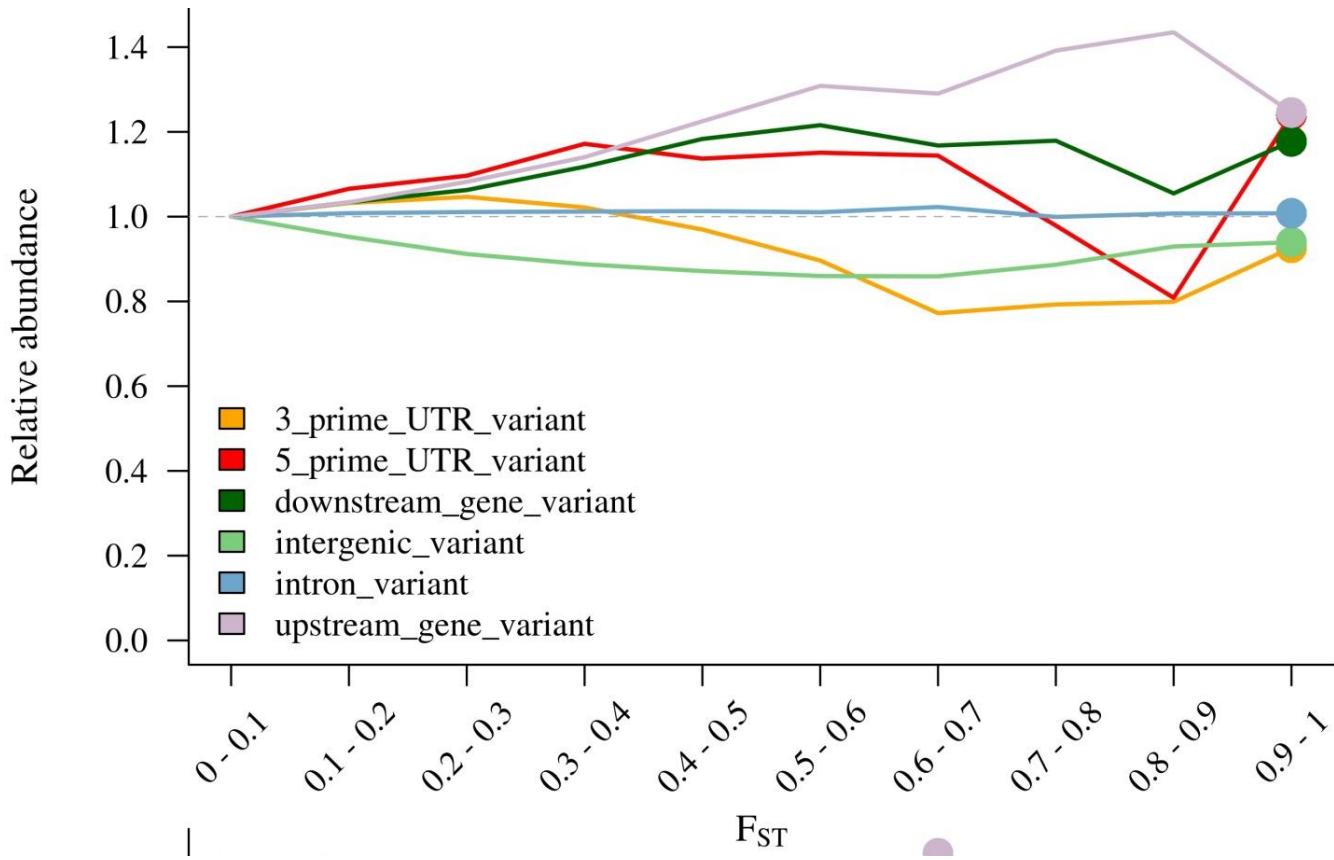
- coding_sequence_variant
- 3_prime_UTR_variant
- intron_variant
- NMD_transcript_variant
- non_coding_transcript_variant
- upstream_gene_variant
- downstream_gene_variant
- intergenic_variant



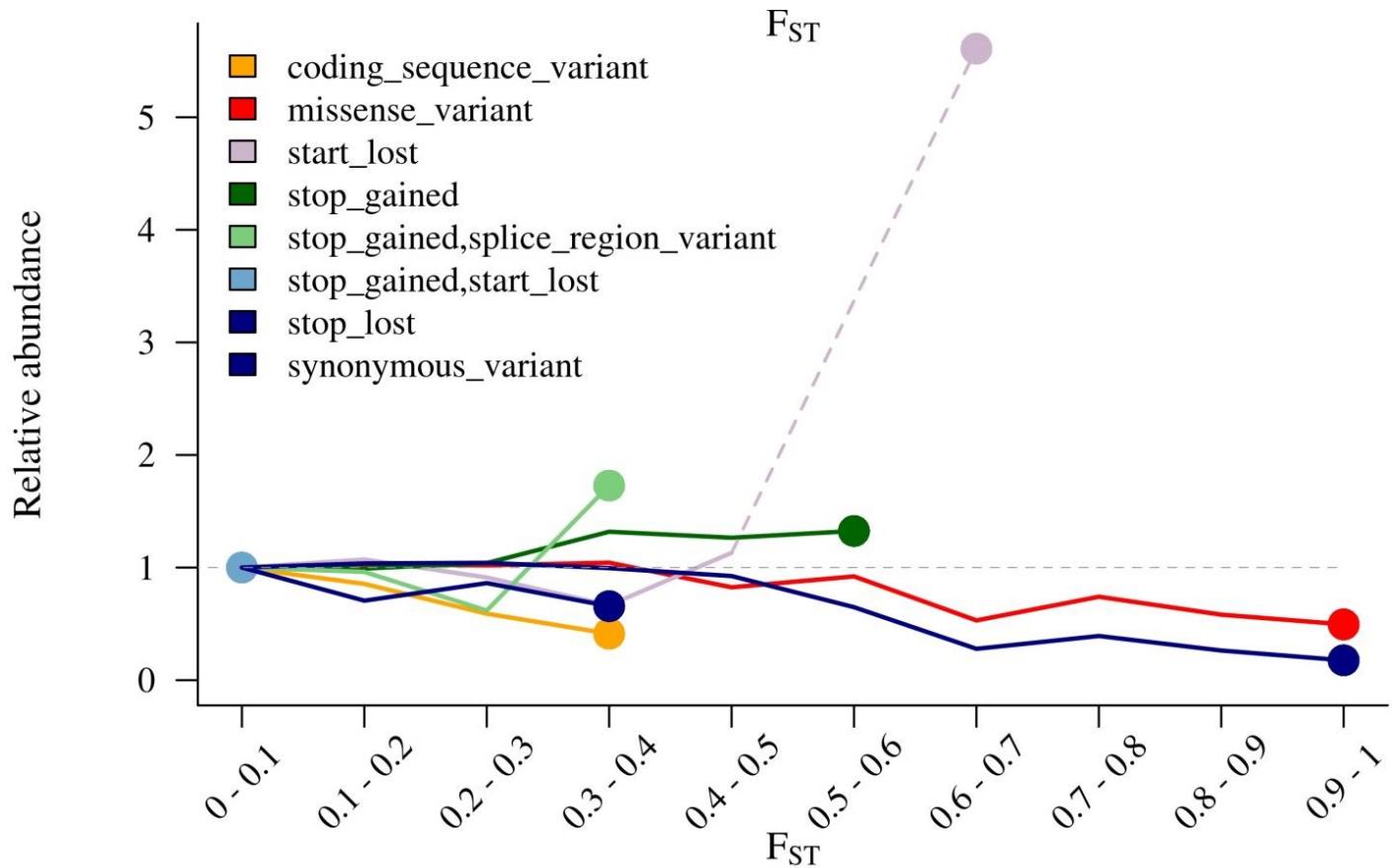
<https://commons.wikimedia.org/w/index.php?curid=5986132>

Consequence type	Count
coding_sequence_variant	4
3_prime_UTR_variant	6
intron_variant	387
NMD_transcript_variant	2
non_coding_transcript_variant	5
upstream_gene_variant	51
downstream_gene_variant	74
intergenic_variant	902

Functional annotation



Functional annotation



Final remarks



- The GMP has a long and stringent breeding history, which resulted in a well-established animal model
- World-wide collaboration of various parties ensures sufficient population size, market access and knowledge gain
- Centralized breeding organization guarantees uniformity of all Göttingen Minipigs, while efficiently maintaining genetic diversity



Thank you!

ELLEGAARD

• •
GÖTTINGEN MINIPIGS



Recognize and
build on your
innovation and
biomedical
differentiation and



Cited and additonal literature



1. Gaerke C, Ytournel F, Sharifi a. R, Pimentel ECG, Ludwig A, Simianer H. 2014. Footprints of recent selection and variability in breed composition in the Göttingen Minipig genome. *Anim Genet* 38:1–391.
2. Fierabracci A. Recent insights into the role and molecular mechanisms of the autoimmune regulator (AIRE) gene in autoimmunity. *Autoimmunity Reviews*. 2011;10:137–43.
3. Hudmon A, Schulman H. Structure-function of the multifunctional Ca²⁺/calmodulin-dependent protein kinase II. *Biochemical Journal*. 2002;364:593–611.
4. Li H, Durbin R. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics*. 2009;25:1754–60.
5. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, et al. The Sequence Alignment/Map format and SAMtools. *Bioinformatics*. 2009;25:2078–9.
6. McLaren W, Gil L, Hunt SE, Riat HS, Ritchie GRS, Thormann A, et al. The Ensembl Variant Effect Predictor. *Genome Biol*. 2016;17:122.
7. Picard. <http://picard.sourceforge.net/>. Accessed 2013-07-26. 2009.
8. Piórkowska K, Żukowski K, Tyra M, Szyndler-Nędza M, Szulc K, Skrzypczak E, et al. The Pituitary Transcriptional Response Related to Feed Conversion in Pigs. *Genes (Basel)*. 2019;10:712.
9. Reimer C, Ha NT, Sharifi AR, Geibel J, Mikkelsen LF, Schlather M, et al. Assessing breed integrity of Göttingen Minipigs. *BMC Genomics*. 2020;21:308.
10. Reimer C, Rubin C-J, Sharifi AR, Ha N-T, Weigend S, Waldmann K-H, et al. Analysis of porcine body size variation using re-sequencing data of miniature and large pigs. *BMC Genomics*. 2018;19:687.
11. Simianer H, Köhn F. Genetic management of the Göttingen Minipig population. *J Pharmacol Toxicol Methods*. 2010;62:221–6.
12. Suzuki T, Minami N, Kono T, Imai H. Zygotically Activated Genes Are Suppressed in Mouse Nuclear Transferred Embryos. *Cloning Stem Cells*. 2006;8:295–304.